

Comparative Immunology of Tuberculosis

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Immune Response - Relevance

1. Diagnostics: Is the animal infected?

2. Vaccination: Did the vaccine elicit a response and is this response protective?

3. Correlations to:

- **Pathology:** response after infection compared to disease
- **Protection:** response after vaccination compared to efficacy
- **Infection:** Active vs latent? Progressive, resolving, or cured?



**Robert Koch,
1890 - develops
tuberculin**

Immune Response – *One Size Does Not Fit All!*

Host Factors:

- Poorly organized vs organized granulomas, badgers vs cattle
- Multibacillary vs paucibacillary disease, mice vs humans/cattle

Pathogen Factors:

- *M. bovis*, *M. tuberculosis*, *M. africanum*, *M. caprae*, *M. pinnipedii*, *M. mungi*, *M. microti*, dassie bacillus, etc
- Hopkins Model - Cavitation in rabbits upon *M. tb* challenge requires prior sensitization to heat killed *M. bovis*

Host / Pathogen Interactions:

- Host adaptation led to speciation
- Speciation partially defined by transmission capacity

Environmental Factors:

- *M. mungi* – prevalence dramatically increases with dry season / Badgers and underground environment
- Nutrition, exposure, over-population, stress, etc.

Louis Pasteur -
1860's, germ
theory of
tuberculosis &
pasteurization



Humans

IFN- γ : humans with defective receptors develop severe disease

TNF- α : essential for proper granuloma formation – TNF inhibitors used for Crohns Disease, rheumatoid arthritis, etc. promote transition from latent to active TB

CD4+ cells: HIV patients more susceptible

IL-12: humans with defective IL12p40 subunit or receptors develop severe disease

Antibody: associated with active disease; sensitivity associated with smear positive cases which correlate with pathology – likely due to antigen load

Latent disease common – latency not documented in other species! Generally only CMI (undetectable antibody) with latent disease.

Chemokines, Cathelicidins, CD8+ cells, IL-17, etc.



Emil von Behring
Nobel Prize, 1901
Also, *M. tb* to cattle

“I need hardly add that the fight against cattle tuberculosis only marks a stage on the road which leads finally to the effective protection of human beings against the disease.”



Laboratory Models of TB

Mice: multibacillary, **high mycobacterial burden** with progressive, less-organized pathology and more neutrophils than many other species

IFN- γ > NO > $\alpha\beta$ TCR > MHC II > MHC I > WT and $\gamma\delta$ TCR - **Th1 immunity crucial**

Kinetics of Infection: **2 wk delay** in activation of T cells, optimal antigen presentation in lymph nodes draining the lungs, then migration of specific T cells back to the lungs

Apoptosis of infected macrophages critical for efficient priming of T cells via uptake of apoptotic vesicles and antigen presentation by dendritic cells - *M. tb* complex mycobacteria actively inhibit apoptosis

Guinea Pigs, Rabbits: pathology similar to humans, models for evaluating DTH and cellular immunity to TB

Skin test in Mice – sensitivity to tuberculin 1000X lower than humans
(*differences exist in application of diagnostic tests*)

Hosts of Veterinary Significance

Cattle: DTH, IFN- γ (Bovigam), poor antibody response,

Cervids: DTH (? Accuracy), moderate antibody response

Eurasian Badgers: Poor DTH response, antibody to MPB83 correlates w/bacterial load & ability to transmit, pathology – poor granuloma, cell-mediated immunity less vigorous yet good enough (low EC response)

Elephants: Robust antibody response, poor DTH (difficult to apply)

South American Camelids: antibody response, poor DTH

Wildboar, Spain: Oral heat-killed *M. bovis* vaccine is effective?

Antigen recognition profiles vary for different species



Correlates of Protection (the Holy Grail)

DTH (skin test) is indicative of prior exposure to *Mycobacteria* spp. but is NOT indicative of disease severity or protection elicited by vaccination

IFN- γ responses, especially to specific antigens such as ESAT-6/CFP10, are indicative of infection but do not necessarily correlate to protection elicited by vaccination

Central Memory Responses (T_{CM}) – correlate to reduced bacterial burden and reduced pathology

IL-17 – correlate to pathology, pre-challenge responses may also correlate to protection.

Multi-functional T cells - IFN- γ TNF- α , IL-2

Patterns of response (multi cytokine / chemokine / etc. profile) – outcome of RNA sequencing studies



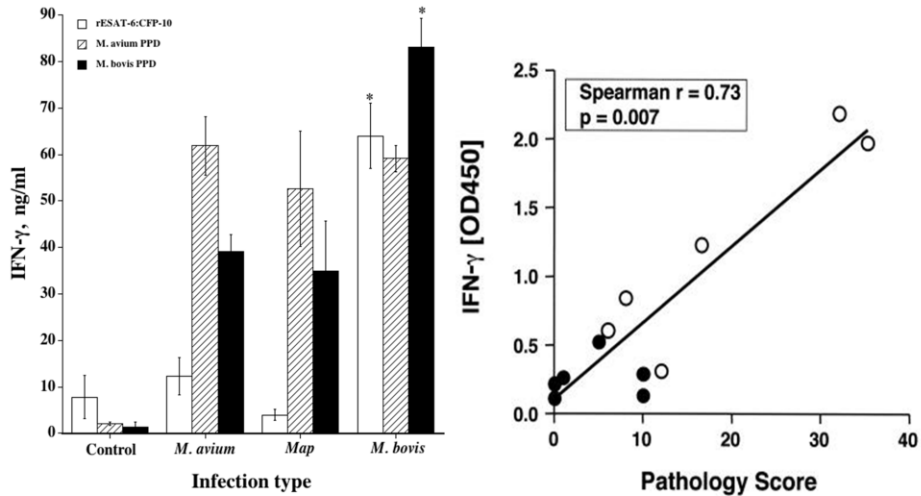
Leonard Pearson
1892 - 1st injection
of tuberculin to US
cattle for TB test

Cattle

IFN- γ Response - Diagnostic and Correlate to Infection

Figure 3

ESAT-6:CFP10 response generally correlates to Pathology



M. avium/Map – PPDa > PPDb, no EC

M. Bovis PPDb > PPDa, +EC

EC response correlates to pathology

Cattle

The IFN- γ Response to PPD_b does not always Correlate to Pathology; however, it is a good Correlate to Infection

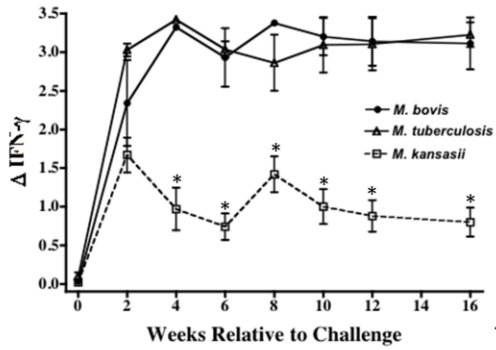
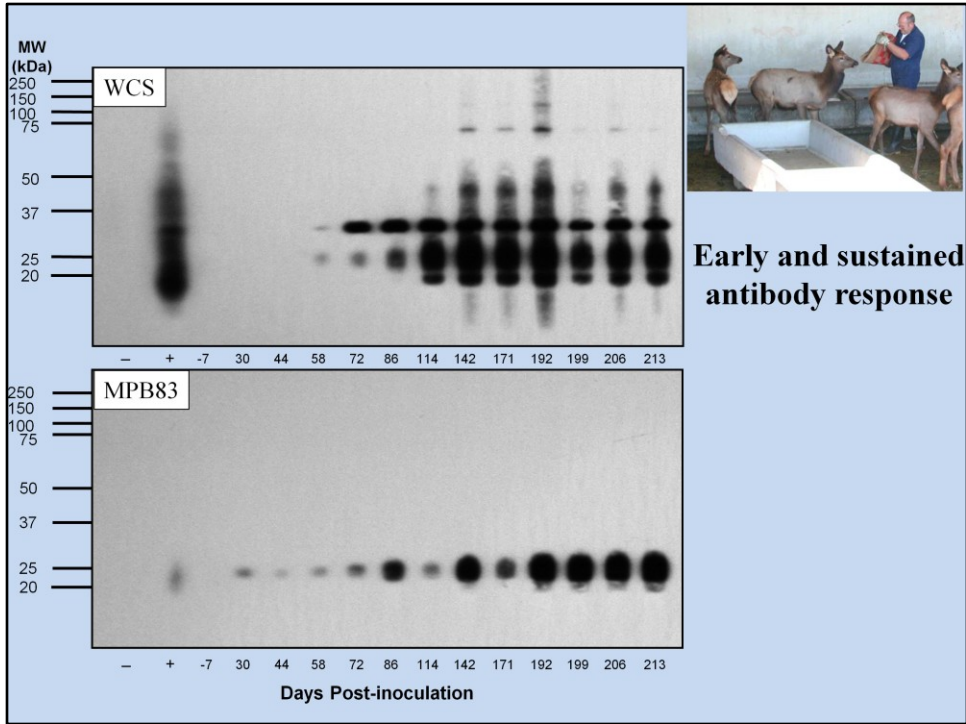
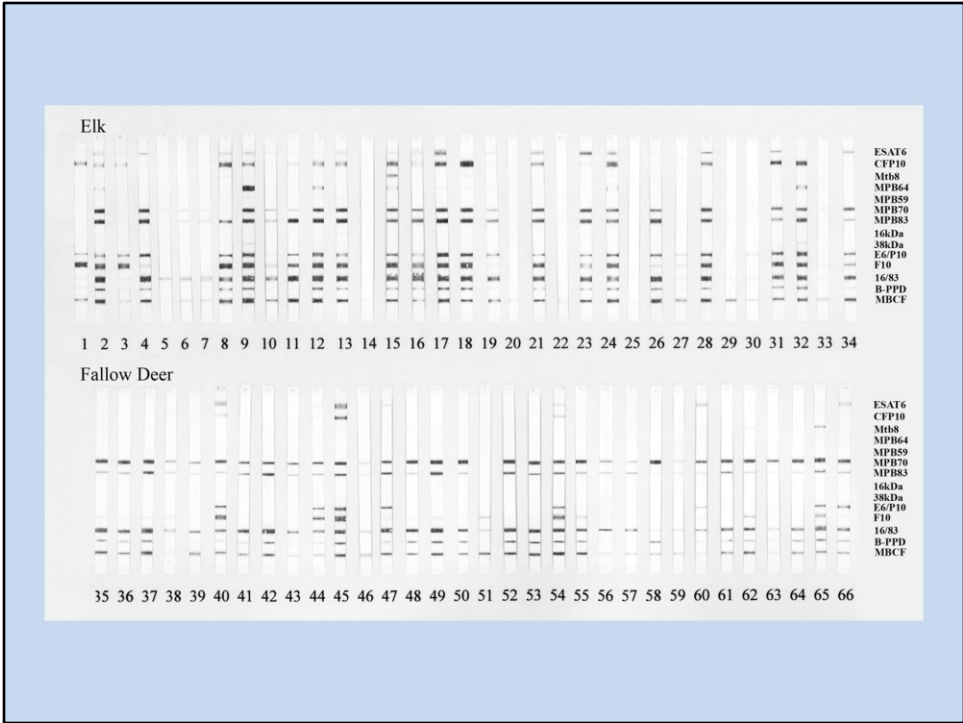


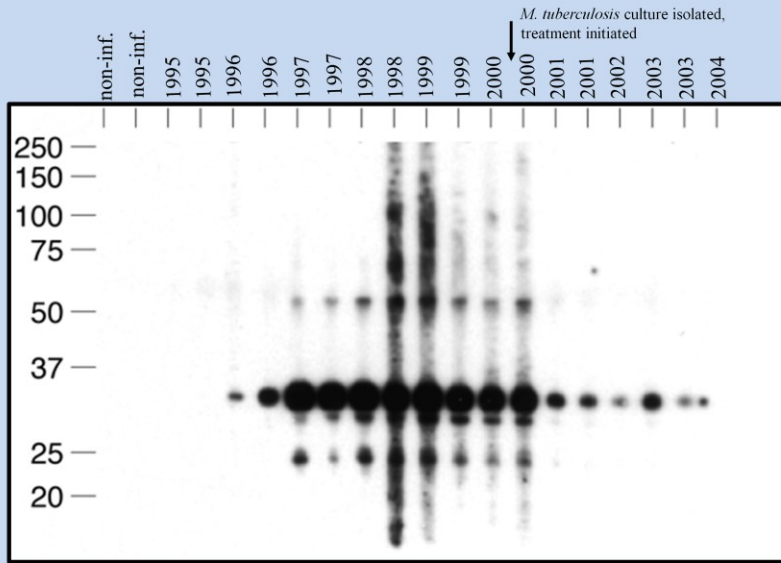
Table 1. Disease expression upon mycobacterial inoculation.

Group	Gross Pathology ^a	Culture ^b *
<i>M. bovis</i> (n = 5)	All positive	27.2 \pm 7.3
<i>M. tuberculosis</i> (n = 5)	All negative	13.9 \pm 5.5
<i>M. kansasii</i> (n = 4)	All negative	0 \pm 0





Immunoblot



Conclusions: Comparative Immunology to TB

- **Similar, yet different, immune responses between hosts / pathogens**

- **Opportunities:**

1. TcM
2. Specific antibody
3. Cytokine profile comparisons
4. Host / pathogen comparisons
5. Interactions
6. Field Application



- **In regards to the immune response to TB, generalizations and extrapolations between hosts is risky!!!**